

Unusual Isoxazoline Formation by Intramolecular Cyclization of (9*E*)-Erythromycin A Oxime

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In the last few years there has been a renewal of interest in macrolide antibiotics, following the successful introduction in the market of second generation derivatives of erythromycin A^{1,2)} with improved pharmacokinetic and antimicrobial activity, like roxythromycin, clarythromycin and azithromycin.

Azithromycin differs structurally from erythromycin A by the insertion of a methyl substituted nitrogen at 9a of the lactone ring, creating a 15-membered macrocycle. This insertion was obtained³⁾ by a Beckmann rearrangement of *E*-oxime **1** in which the nitrilium ion intermediate was trapped intramolecularly by the 6-oxygen giving rise to the imino ether **2** that was reduced subsequently to the amine **3** and finally methylated to **4**.

When the Beckmann rearrangement is performed in ether⁴⁾, in the presence of pyridine and with addition of *p*-toluensulphonyl chloride at -45°C , the nitrilium ion is trapped also by the O-11, giving a mixture of **2** and **5**. Carrying out the reaction in the same conditions at

$0\sim 5^{\circ}\text{C}$ the lactam **6** can be isolated³⁾ as the major product. Similar compounds with the annular nitrogen at 8a of the macrolide ring were synthesised by the Beckmann rearrangement of the *Z*-erythromycin oxime, which is readily available by the basic isomerization of the corresponding *E* isomer⁵⁾.

During our work on erythromycin derivatives we unexpectedly isolated isoxazoline **7** in conditions very similar to those required for the preparation of imino ether **2**³⁾ but the choice of base. In fact **2** is obtained by DJOKIC³⁾ in 86.5% yield using excess of sodium hydrogen carbonate and tosyl chloride in a 1:1 water-acetone mixture. If at variance with respect to the conditions reported above the *E* oxime **1** is treated with tosyl chloride in a 1:5 water-acetone mixture with excess potassium hydroxide as a base we obtained **7** in 60% yield. The structure of **7** was determined on the basis of ¹H, ¹³C and ¹⁵N NMR studies. The ¹⁵N spectra were run on a concentrated solution of **7** in CDCl₃ (ca. 300 mg/0.5 ml) under proton broad-band decoupling and with the suppression of the heteronuclear NOE (the ¹⁵N nucleus has negative ¹H-¹⁵N nuclear Overhauser effect leading to a decrease in signal intensity or even to a complete disappearance of the signal). The ¹⁵N spectrum of **7** shows two peaks at -361 and -24 ppm relative to the external neat nitromethane. The resonance at -361 ppm is characteristic⁶⁾ of amino nitrogen atoms and can be assigned to the N(CH₃)₂ group of the desosamine residue. The resonance at -24 ppm cannot be attributed to any structure deriving from the Beckmann rearrangement; this rearrangement, as mentioned before, should afford some imino ether (6,9- or 9,11-imino ethers, ¹⁵N region $-130\sim -170$ ppm⁶⁾) or lactamic derivatives (¹⁵N region ca. -250 ppm⁶⁾) de-

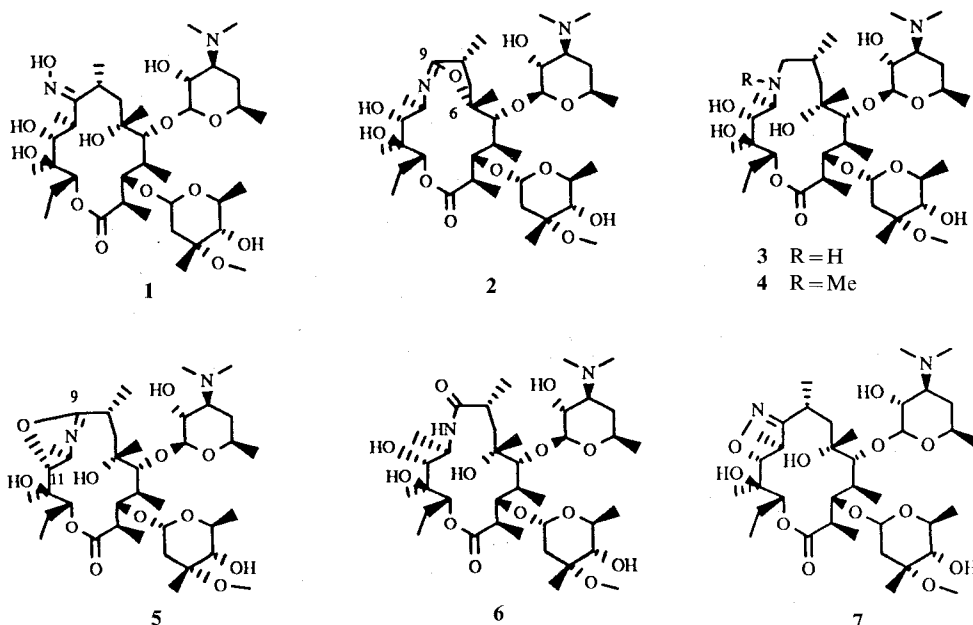


Fig. 1.

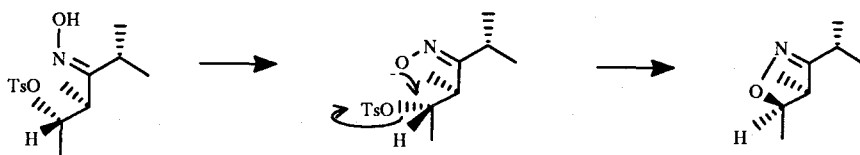
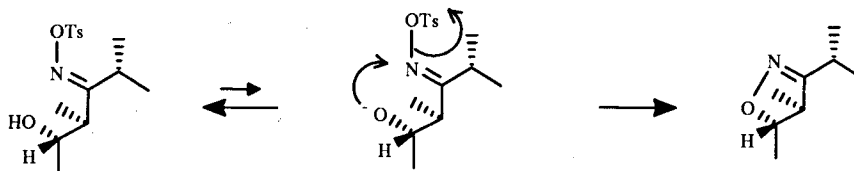


Fig. 2.



pending on which hydroxyl group is captured by the intermediate nitrilium structure³). The observed resonance at -24 ppm falls in the region of an oximic and isoxazolinic nitrogen atom (-20 – -45 ppm^{6,7}) and is in agreement with our structure.

The formation of **7** from **1** can be explained by, at least, two different mechanisms (Figs. 1 and 2).

The first requires the isomerization of the *E* oxime to *Z* oxime, followed by tosylation of O-11 with subsequent nucleophilic substitution on C-11 (Fig. 1). The second mechanism, simply envisions the tosylation of the *E* oxime oxygen, the normal pathway in the Beckmann formation of iminoether **5**, followed by a direct nucleophilic attack of the 11 hydroxyl, eventually in an ionised form, onto the oximic nitrogen with prior or concomitant release of tosylate anion (Fig. 2).

The second mechanism seems more likely to us on the basis of the following observations: i) the *E* oxime **1** dissolved in acetone at 0°C in the presence of KOH does not show isomerization to the *Z* oxime for at least 5 hours later; on the contrary the isomerization occurs at room temperature but the process is slow and takes some hours⁵ to reach the equilibrium; ii) the configuration of carbon C-11 of the isoxazoline derivative **7** is the same as the starting *E* oxime **1**. While the vicinal coupling constant $J(10, 11)$ of 8.7 Hz is not conclusive for the elucidation of the relative stereochemistry of carbons C-10 and C-11, the large NOE enhancement observed between H-10 and H-11 (*ca.* 9%) and the absence of NOE between H-11 and CH₃-10 strongly supports the assumption that hydrogens H-10 and H-11 are located *cis* on the isoxazoline ring. The lack of inversion of C-11 configuration rules out the S_N2 type reaction suggested by the first mechanism. The second mechanism, leading to the formation of an isoxazoline ring is, as far as we know, very unusual. On the other hand this mechanism has been invoked previously⁸) to explain the formation under the conditions of the Beckmann rearrangement of

an isoxazoline ring in a steroidal scaffold probably due to conformational biases. It is reasonable to assume the same should be true also for our macrolide isoxazoline. In fact detailed conformational studies carried out on erythromycin oxime⁹) and a derivative (*E*)-11-*O*-(2-dimethylaminoethoxy)methyl-9-deoxy-9-methoxyiminoerythromycin A¹⁰) indicated that the C₉=N₉ bond is almost parallel to the C₁₁–O₁₁ bond (the dihedral angle O₁₁–C₁₁–C₉–N₉ is less than 5°) and the distance O₁₁⋯N₉ is about 3 \AA , approximately the sum of the Van der Waals radii of the oxygen and nitrogen atoms. Such geometry together with the strong basic conditions of the reaction should favour formation of the oxygen-nitrogen bond instead of the usual Beckmann rearrangement.

Compound **7** shows extraordinary acid stability ($t_{1/2} = 24$ hours at pH 1) even in comparison with second generation macrolide antibiotics that were developed for their resistance to acidic decomposition, typical of this class of compounds. This stability could be of conformational origin and a study on the subject will be reported elsewhere.

Experimental

A solution of 10 g (13.37 mmol) of *E*-erythromycin oxime in 200 ml of acetone at 0°C was treated with 53.48 mmol of KOH dissolved in 50 ml of water. After 5 minutes, 26.74 mmol of TsCl dissolved in 50 ml of acetone were added. After 1.5 hours at 0°C and 1.5 hours at room temperature the acetone was evaporated *in vacuo* and the residue partitioned between CH₂Cl₂ and water. The organic phase was separated, dried over Na₂SO₄ and evaporated *in vacuo*. The crude product was purified by Flash Chromatography (CH₂Cl₂:MeOH:NH₄OH 90:10:1) giving **7** in 60% yield. MS (EI): 730. mp 253°C . $\alpha_D = -125^\circ$ ($c = 1$ in CH₂Cl₂).

NMR Data for 7

^1H NMR (CDCl_3), δ 2.97 (dq, 1H, H-2, $J=3.9$, 7.4 Hz), 4.14 (dd, 1H, H-3, $J=1.8$, 3.8 Hz), 2.74 (m, H-4, $J=1.5$, 7.5 Hz), 3.54 (d, 1 H, H-5, $J=8.2$ Hz), 1.39 (d, 1H, H-7, $J_{\text{gem}}=13.6$ Hz), 2.78 (dd, 1H, H-7', $J=11.1$, 13.6 Hz), 2.59 (m, 1 H, H-8, $J=11.1$, 7.1, 1.0 Hz), 3.59 (dq, 1H, H-10, $J=7.5$, 8.6 Hz), 4.45 (d, 1H, H-11, $J=8.6$ Hz), 4.65 (dd, 1H, H-13, $J=1.9$, 11.2 Hz), 1.61 (m, 1H, H-14, $J_{\text{gem}}=14.0$ Hz, $J=7.2$, 2.0 Hz), 1.93 (m, 1H, H-14', $J_{\text{gem}}=14.0$ Hz, $J=7.2$, 11.4 Hz), 0.91 (t, 3H, CH_3 -14, $J=7.1$ Hz), 1.13 (d, 3H, CH_3 -2, $J=7.4$ Hz), 1.10 (d, 3H, CH_3 -4, $J=7.5$ Hz), 1.24 (s, 3H, CH_3 -6), 1.08 (d, 3H, CH_3 -8, $J=7.3$ Hz), 1.51 (d, 3H, CH_3 -10, $J=7.5$ Hz), 1.14 (s, 3H, CH_3 -12), 4.37 (d, 1H, H-1', $J=7.3$ Hz), 3.23 (dd, 1H, H-2', $J=10.1$, 7.4 Hz), 2.46 (m, 1H, H-3', $J=10.2$, 3.9, 12.3 Hz), 1.22 (m, 1 H, H-4'ax), 1.68 (m, 1H, H-4'eq, $J=1.8$, 3.9, 14.4 Hz), 3.46 (dq, 1H, H-5', $J=1.8$, 6.1, 10.6 Hz), 1.22 (d, 3H, CH_3 -5', $J=6.1$ Hz), 2.31 (s, 6H, $\text{N}(\text{CH}_3)_2$), 1.56 (dd, 1H, H-2''ax, $J=15.2$, 4.8 Hz), 2.41 (d, 1H, H-2''eq, $J=15.2$ Hz), 3.02 (dd, 1H, H-4'', $J=9.6$, 10.5 Hz), 4.03 (dq, 1H, H-5'', $J=9.2$, 6.2 Hz), 3.34 (s, 3H, OCH_3 -3''), 1.24 (s, 3H, CH_3 -3''), 1.27 (d, 3H, CH_3 -5'', $J=6.2$ Hz), 2.18 (d, 1H, OH-4'', $J=10.5$ Hz).

^{13}C NMR (CDCl_3), δ 178.8 (C-1), 44.9 (C-2), 78.6 (C-3), 42.7 (C-4), 82.4 (C-5), 37.4 (C-7), 26.2 (C-8), 168.3 (C-9), 45.3 (C-10), 85.1 (C-11), 82.6 (C-13), 23.0 (C-14), 11.4 (CH_3 -14), 103.4 (C-1'), 70.8 (C-2'), 65.7 (C-3'), 29.1 (C-4'), 68.9 (C-5'), 21.5 (CH_3 -5'), 40.4 ($\text{N}(\text{CH}_3)_2$), 95.4 ((C-1''), 34.7 (C-2''), 78.0 (C-4''), 65.5 (C-5''), 18.4 (CH_3 -5''), 49.6 (OCH_3 -3''), 21.4 (CH_3 -3''), 13.6 (CH_3 -2), 8.9 (CH_3 -4), 29.3 (CH_3 -6), 20.4 (CH_3 -8), 10.0 (CH_3 -10), 23.5 (CH_3 -12), 75.8 and 72.9 (signals due to the three quaternary carbons C-12, C-6 and C3'', which were not assigned).

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